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A comparative evaluation of coenzyme Q10-loaded liposomes and solid lipid nanoparticles as dermal antioxidant carriers

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Background: The effective delivery of coenzyme Q10 (Q10) to the skin has several benefits in therapy for different skin pathologies. However, the delivery of Q10 to deeper layers of skin is challenging due to low aqueous solubility of Q10. Liposomes and solid lipid nanoparticles (SLN) have many advantages to accomplish the requirements in topical drug delivery. This study aims to evaluate the influence of these nanosystems on the effective delivery of Q10 into the skin.

Methods: Q10-loaded liposomes (LIPO-Q10) and SLNs (SLN-Q10) were prepared by thin film hydration and high shear homogenization methods, respectively. Particle size (PS), polydispersity index (PI), zeta potential (ZP), and drug entrapment efficiency were determined. Differential scanning calorimetry analysis and morphological transmission electron microscopy (TEM) examination were conducted. Biocompatibility/cytotoxicity studies of Q10-loaded nanosystems were performed by means of cell culture (human fibroblasts) under oxidative conditions. The protective effect of formulations against production of reactive oxygen species were comparatively evaluated by cytofluorometry studies.

Results: PS of uniform SLN-Q10 and LIPO-Q10 were determined as 152.4 ± 7.9 nm and 301.1 ± 8.2 nm, respectively. ZPs were -13.67 ± 1.32 mV and -36.6 ± 0.85 mV in the same order. The drug entrapment efficiency was 15% higher in SLN systems. TEM studies confirmed the colloidal size. SLN-Q10 and LIPO-Q10 showed biocompatibility towards fibroblasts up to 50 μ M of Q10, which was determined as suitable for cell proliferation. The mean fluorescence intensity % depending on ROS production determined in cytofluorometric studies could be listed as $Q10 \cong SLN-Q10 > LIPO-Q10$.

Conclusion: The LIPO-Q10 system was able to enhance cell proliferation. On the contrary, SLN-Q10 did not show protective effects against ROS accumulation. As a conclusion, liposomes seem to have advantages over SLN in terms of effective delivery of Q10 to skin for antioxidant purposes.

Keywords: solid lipid nanoparticles, liposomes, coenzyme Q10, antioxidant, cytotoxicity

Introduction

The skin is the body's largest organ with an area of approximately 2 m². Its main function is to protect the body from external threats such as toxins, pathogens, and UV radiation. After being absorbed by molecules in the skin, UVA initiates the formation of reactive oxygen species (ROS), causing oxidative damage to lipids, proteins, and DNA.¹ Several endogenous mechanisms also contribute to the production of ROS and excessive amounts of ROS is thought to be a key contributor to some pathologies like cancers.^{2,3} To be able to cope with these oxidative stresses produced by internal and external factors, the skin has both enzymatic (superoxide dismutase, catalase,

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and glutathione peroxidase) and nonenzymatic (antioxidant) mechanisms for protection.¹

Q10, the only lipid soluble endogenous antioxidant, is a component of the electron transport chain in most eukaryotic cells, primarily in the mitochondria. It has an important role in generating adenosine triphosphate during aerobic cellular respiration and inhibits cell membrane peroxidation in the reduced form.^{4,5} However, to deliver Q10 to deeper layers of skin is challenging due to the low aqueous solubility of Q10 and due to the barrier function of stratum corneum.⁶

As lipidic nanocarriers, liposomes and solid lipid nanoparticles (SLN) have taken the interest of scientists for their potential to overcome the topical drug delivery challenges.⁷ Liposomes are promising delivery systems that accomplish the requirements for a topical drug delivery. Topical liposomes may increase solubilization and act as a local depot for sustained release of dermally active compounds, and can enhance penetration.⁸ SLNs are alternative colloidal carrier systems based on solid lipids. SLNs for dermal applications include many advantages such as occlusion, drug targeting, and modulation of drug release.⁹ SLNs favor drug penetration into the skin and sustain the release to avoid systemic absorption.¹⁰ Chemically unstable compounds that are sensitive to light, oxidation, and hydrolysis can be protected in the form of SLNs.^{11,12}

In light of this knowledge, this study was designed to evaluate the influence of these two lipid based nanosystems on the effective delivery of Q10 into the skin. Q10-loaded liposomes (LIPO-Q10) and SLNs were prepared, characterized, and their biocompatibility/cytotoxicity was evaluated by means of cell culture (human fibroblasts) under oxidative conditions. Their antioxidant effects on the production of ROS were comparatively evaluated by cytofluorometry studies.

Materials and methods

Materials

Phosphatidylcholine from soybean (Lipoid S100) was supplied by Lipoid (Ludwigshafen, Germany) and Labrasol® Compritol® 888 ATO (glyceryl behenate) was obtained from Gattefossé (Lyon, France). Poloxamer® 188 (Pluronic® F68) was donated by BASF (Ludwigshafen, Germany). Cholesterol, Tween® 80 (polysorbate 80) and Q10 were obtained from Sigma-Aldrich (St Louis, MO). All high-performance liquid chromatography (HPLC) reagents and chloroform were purchased from Sigma-Aldrich. The other chemicals were obtained from Merck KGaA (Darmstadt, Germany). All filters were purchased from Sartorius (Göttingen, Germany).

Preparation of liposomes

The thin film hydration method was used to prepare the liposomal suspensions.¹³ Phospholipid-Lipoid S100 (25 mg), cholesterol (6.25 mg), and Q10 (5 mg) were dissolved in chloroform (12.5 mL), and organic solvent was evaporated at 40°C to form a thin film (Ika RV 10 rotary evaporator; IKA®-Werke GmbH & Co., Staufen, Germany). Subsequently, the resulting thin film was hydrated in 25 mL of bidistilled water by rotating in the same evaporator at 100 rpm without using any vacuum. The liposomal suspension was homogenized using ULTRA-TURRAX® (IKA® T25; IKA®-Werke) at 3500 rpm.

Preparation of SLNs

Q10-loaded SLNs (SLN-Q10) were prepared by the high shear homogenization method.¹⁴ C888 was employed as the lipid base, Poloxamer 188 (P188) and Tween 80 (Tw 80) were used as surfactant and cosurfactant, respectively. As reported in our previous study,¹⁴ the lipid phase consisting of 300 mg Compritol® 888 ATO (C888) and 5 mg Q10 were mixed and heated to 85°C. At the same time, the aqueous phase consisting of 150 mg of P188 and 75 mg of Tw 80 in 12.5 mL bidistilled water was heated to the same temperature. The aqueous phase was poured into the lipid phase drop by drop and mixed with ULTRA-TURRAX (T25) at a speed of 24,000 rpm. Then the pre-emulsion produced was dispersed in 12.5 mL bidistilled water at 4°C and kept at -20°C for 10 minutes to obtain nanoparticles by solidification. Blank SLNs were prepared in a similar way, without addition of Q10.

Measurement of particle size (PS) and polydispersity index (PI)

Liposomes and SLN dispersions were characterized in terms of PS and PI at 25°C by photon correlation spectroscopy (Zetasizer-Nano ZS; Malvern Instruments Ltd, Malvern, UK) at an angle of 173°. The formulations were diluted with bidistilled and filtered (0.45 µm) water before the experiment (n = 6). Each sample was measured in triplicate.

Measurement of zeta potential (ZP)

The ZP of liposomes and SLN dispersions was measured at 25°C, under an electrical field of 40 V/cm (Zetasizer-Nano ZS). The measurements were conducted in triplicate.

Drug entrapment efficiency

SLN dispersions or liposomes were placed in a dialysis bags (cutoff 25 kDa). These bags were centrifuged for 1 hour at

14,000 rpm in a medium consisting of 5% of Labrasol. Then the amount of Q10 in 5% Labrasol solution was analyzed by a validated HPLC method and the quantity of free drug was calculated. The encapsulated amount of Q10 was calculated by subtracting the free amount of Q10 from the total amount in the dispersion. Each batch was evaluated three times. The following equation was used to calculate the entrapment efficiency (EE%), where W_i is the amount of initial drug and W_f is the amount of free drug:

$$EE\% = \frac{W_i - W_f}{W_i} \times 100$$

The HPLC system (Agilent Series 1100) consisted of a C18 reverse phase column (ACE 5-C18 250 mm × 4.6 mm). The mobile phase was a mixture of 65:35 methanol:*n*-hexan (v/v).¹⁵ The flow rate and UV wavelength were set at 1 mL/minute and 275 nm, respectively.

TEM (transmission electron microscopy) analysis

TEM (CM12 Philips; Philips, Amsterdam, The Netherlands) was used for the morphological examination of liposomes and SLNs. 2% (w/v) phosphotungstic acid was used to stain the samples and after staining they were placed on copper grids for viewing.

Differential scanning calorimetry (DSC) analysis

Q10, C888, a physical mixture of C888:Q10, blank/loaded SLN, and blank/loaded liposome formulations were analyzed. The samples to be tested were sealed in aluminum pans (50 μL) and placed in differential scanning calorimeter (DSC 8000; PerkinElmer, Waltham, MA). DSC analysis was conducted under nitrogen flow (20 mL/minute) in a temperature range between 30°C–300°C.

Cell culture studies

Cytotoxicity studies

Cells between the second and fifth passage of normal human dermal fibroblasts from juvenile foreskin (PromoCell GmbH, Heidelberg, Germany) were employed for cytotoxicity studies. The growth media of fibroblasts consisted of Dulbecco's Modified Eagle Medium (DMEM; Sigma-Aldrich) augmented with 10% foetal bovine serum (Sigma-Aldrich) with 200 IU/mL penicillin, and with 0.2 mg/mL streptomycin. The cells were kept in an incubator with an atmosphere of 5% CO₂ and 95% relative humidity at 37°C.

Ninety-six-well plates (area of 0.34 cm²) were used to seed the fibroblasts at a density 10⁵ cells/cm² in each well. Cells were grown for 24 hours to obtain sub-confluence. After 24 hours the medium was removed and the samples to be tested were replaced. The cells were put in contact with 200 μL of SLN-Q10 and LIPO-Q10 in the concentration range of 10–250 μM. After the incubation of cell substrates for 24 hours, the medium was removed and the (3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) (MTT) test was performed. 125 μL of MTT solution (Sigma-Aldrich) at 0.25 μg/mL in Hank's buffered salt solution (pH 7.4) was put in contact with each cell substrate for 3 hours. An enzyme-linked immunosorbent assay plate reader (iMark[®] absorbance reader; Bio-Rad, Hercules, CA) at a wavelength of 570 nm with a reference wavelength of 655 nm was used to assay the absorbance. The % ratio between the absorbance of each sample and the absorbance of cell substrate maintained in contact with the growth medium was expressed as cell viability.¹⁰

Proliferation studies

To evaluate the effect of oxidative stress on cell viability, additional cell culture studies were seeded in each well of 96-well plates and grown in the same conditions as for cytotoxicity studies. After 24 hours, the media in the wells were replaced with SLN and liposomes at two selected Q10 concentrations (25 μM and 50 μM). After 4 hours of contact, 1.5 mM H₂O₂ was added in these wells. This was the optimum H₂O₂ concentration causing oxidative damage avoiding cell death. After an additional 24 hours of contact, the viability was evaluated with an MTT test and the cell viability was determined as previously described.¹⁰

Evaluation of intracellular ROS accumulation

2,7-dichlorofluoresceine (DCFH-DA) acetate was employed as the fluorescent probe to evaluate intracellular accumulation of ROS. In this assay, cells are incubated with DCFH-DA and are able to cross the cell membrane. Once inside, the DCFH-DA loses its acetate groups and is no more able to go out of the cell. At this point it becomes nonfluorescent (DCFH). Only in the presence of ROS can it be oxidized in DCF, becoming fluorescent again. Higher fluorescence intensity means higher oxidative damage and then lower antioxidant activity.

Fibroblasts were seeded in 12-well plates (area of 3.8 cm²) at a density 10⁵ cells/cm². Cells were grown for 48 hours to obtain subconfluence. 50 μM of SLN-Q10, LIPO-Q10, and Q10 alone, were put in contact with the cells. 1.5 mM H₂O₂ was added in each well after 4 hours of contact. Cell

substrates were incubated for a further 24 hours. Subsequently 1 mM DCFH-DA was put in cell substrates for 15 minutes. To obtain a suspension, cells were scraped from the bottom of each well. This suspension was centrifuged at 1500 rpm for 5 minutes. After the removal of supernatant, the cell pellet was re-suspended in 500 μ L phosphate buffered solution. A cytofluorometer (Navios Flow Cytometer; Beckman Coulter, Inc, Brea, CA) was used to assay these cell suspensions at the excitation wavelength of 488 nm and the emission wavelength of 525 nm. The data were analyzed by Kaluza Analysis software (Beckman Coulter Inc).¹⁶

Statistical analysis

Statistical analysis was conducted by ANOVA followed by Tukey's test for comparisons between groups. The significance level was taken as 95% ($P < 0.05$).

Results and discussion

The physical properties of nanosystems might affect in vivo behavior. In our previous study, it was reported that PS and ZP have important roles in penetration properties of lipid nanoparticles.⁹ The data in terms of PS, PI, ZP, and EE% of SLNs and liposomes prepared with and without Q10 incorporation are shown in Table 1.

The PS obtained with SLNs, in either a blank or loaded state, were significantly smaller than the liposome formulations. The Q10 loading did not affect the PS of SLN system significantly. However, the addition of Q10 decreased the dimension of liposomes ($P < 0.05$). It is known that lipid composition has a significant impact on liposome size and drug loading.¹⁷ A decrement in the liposome dimension might be due to the shrinkage of the aqueous volume of the vesicle resulting in smaller dimensions by the addition of Q10, which is a very lipophilic substance ($\log P \gg 10$).¹⁸ This phenomenon could be explained by either the osmotic effect or the membrane folding. Since the total area of the membrane of each liposome was maintained, the folding of the membrane might cause this size reduction. In a study by Nomura et al¹⁹ the liposomes gradually shrank in size

(5 times smaller) with the addition of peptides and the folding/piling up of liposomal membranes was confirmed by electron microscopy observations.

PI values of the formulations indicated that uniform colloidal systems could be formed for both liposomes and SLNs, with the methods applied in this study. Q10 addition reduced the overall charge of the SLNs as it was seen also in our previous studies conducted with resveratrol and cyclosporine A.^{10,14} On the contrary, Q10 addition increased the total electrical charge of liposomes. Similarly, in a study by Jukanti et al the electrical charge significantly increased from -34.1 ± 2.6 mV up to -46.9 ± 2.7 mV in case of Q10 addition in liposomes.²⁰

The drug entrapment efficiency of Q10 was determined as 73.1% and 89.2% for liposomes and SLNs respectively, by a validated HPLC method. The enhanced encapsulation efficiency determined with SLNs in comparison to liposomes was an expected result, since it is one of the advantages of SLN over liposomes. Besides, the EE% of Q10 in liposomes was also enough to say that Q10 could be encapsulated in liposomes successfully by film hydration method. Verma et al reported that film hydration was the most efficient method for Q10 encapsulation into liposomes in comparison to other methods.²¹ It is known that the partition of the drug between the lipid of membrane and the water, influences the EE% of drugs in liposomes.²² Since Q10 is very lipophilic, its partition would be in favor of hydrophobic regions. Therefore, it was considered that Q10 might have been trapped in the double layers of phospholipids. TEM images acted as evidence for the colloidal size and homogenous structure of liposomes and SLNs (Figure 1).

DSC evaluation can give good clues about whether a drug is entrapped in the bilayer or in the aqueous compartment of the liposomes²³ and helps to understand the melting and re-crystallization behavior of materials like lipid nanoparticles.²⁴ DSC analysis was performed to examine the melting and crystallization properties of SLN and liposomes. The DSC thermograms of the formulations were given in Figures 2 and 3.

Table 1 PS, PI, ZP, and EE% of blank and Q10-loaded SLNs and liposomes

Formulation	PS \pm SD (nm)	PI \pm SD	ZP \pm SD (mV)	EE% \pm SD
Blank liposome	367.9 \pm 7.5	0.413 \pm 0.01	-32.8 \pm 0.2	-
LIPO-Q10	301.1 \pm 8.2	0.458 \pm 0.03	-36.6 \pm 0.9	73.1 \pm 1.7
Blank SLN	164.1 \pm 9.4	0.294 \pm 0.05	-18.64 \pm 1.2	-
SLN-Q10	152.4 \pm 7.9	0.272 \pm 0.03	-13.67 \pm 1.3	89.2 \pm 3.8

Note: The results are the means \pm SD (n = 6).

Abbreviations: PS, particle size; PI, polydispersity index; ZP, zeta potential; EE%, entrapment efficiency; Q10, coenzyme Q10; SLN, solid lipid nanoparticle; SD, standard deviation; LIPO-Q10, Q10-loaded liposomes; SLN-Q10, Q10-loaded solid lipid nanoparticles.

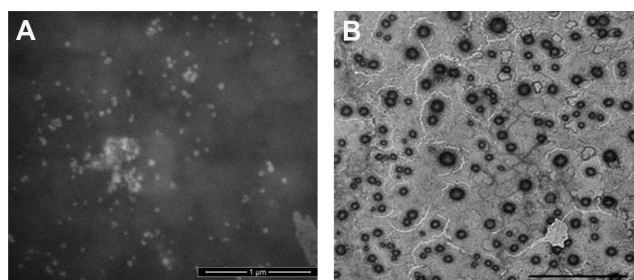


Figure 1 TEM images of (A) SLN-Q10 and (B) LIPO-Q10.

Notes: (A) The bar is 1 μm . (B) The bar is 5 μm .

Abbreviations: TEM, transmission electron microscopy; SLN-Q10, Q10-loaded solid lipid nanoparticles; LIPO-Q10, Q10-loaded liposomes.

The melting points for C888 and Q10 were determined as 72.31°C and 53.96°C, respectively. It was observed that C888 was crystallized during the cooling process of SLN production (Figure 2). The melting peak of Q10 could not be observed in SLN formulation, due to its solubility in the lipid phase (Figure 2). In the DSC thermogram of Q10-loaded liposomal dispersion (Figure 3), the Q10 peak was also lost, indicating good interaction of all components. Since Q10 is highly lipophilic it is possible that it might be entrapped in the bilayer compartment of liposome.

It is known that the structure of liposomes and SLNs may be destroyed by several enzymatic mechanisms such as lipases.¹⁴ However, these systems protect labile drugs from degradation by encapsulation and increase their permeation. Zhou et al²⁵ prepared Q10-loaded lipid nanocapsules with Nile red and observed the accumulation through skin layers by the intensity of fluorescence. It was determined that lipid nanocapsules could be targeted to both epidermis and dermis. In addition, the *in vitro* assessment of the cytotoxicity on human dermal fibroblasts has been used as an approach to evaluate the toxicity of the different formulations.²⁶

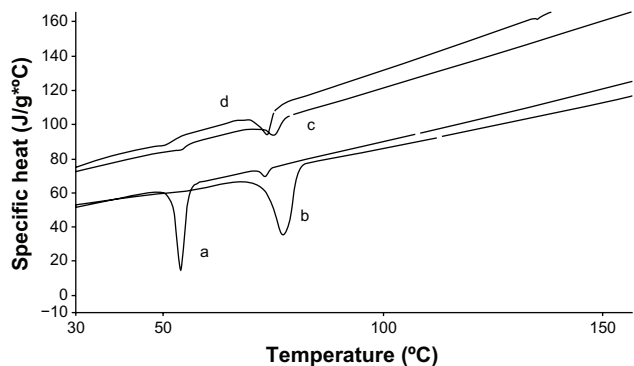


Figure 2 DSC thermograms of (a) Q10; (b) Compritol; (c) blank SLN; and (d) SLN-Q10.

Abbreviations: DSC, differential scanning calorimetry; Q10, coenzyme Q10; SLN, solid lipid nanoparticle; SLN-Q10, Q10-loaded solid lipid nanoparticles.

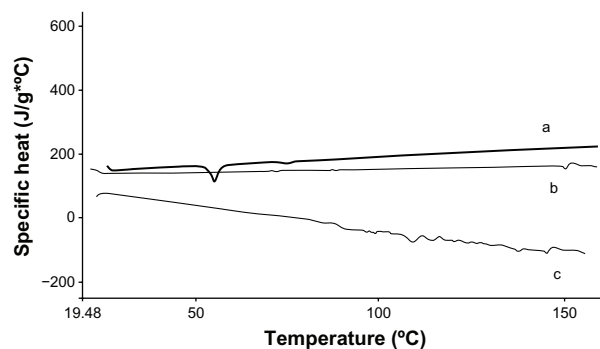


Figure 3 DSC thermograms of (a) Q10 and (b) blank liposome and (c) LIPO-Q10. **Abbreviations:** DSC, differential scanning calorimetry; Q10, coenzyme Q10; LIPO-Q10, Q10-loaded liposomes.

The delivery of Q10 to fibroblasts is particularly important because oxidative stress affects several pathways especially in dermal fibroblasts. A major cause of oxidative stress in skin is solar UV radiation. This phenomenon influences pathways in ways that closely mimic ROS.²⁷ Recently, Reelfs et al have shown that labile iron in skin fibroblast, which is involved in the activation of a number of transcription factors, was released by UVA irradiation.²⁸ It is also known that $^1\text{O}_2$ can initiate c-jun N-terminal kinase signaling, which leads to interstitial collagenase induction as well as the synthesis of proinflammatory cytokines in UVA-irradiated fibroblasts.²⁷ It was also proven that aging skin fibroblasts have decreased catalase activity. Accumulation of ROS owing to catalase attenuation may be a critical aspect of the MAPK signaling changes that result in skin aging and photoaging in human skin *in vivo*.²⁹ Therefore, it was thought that evaluation of the effective delivery of Q10 in dermal fibroblasts will be beneficial due to its involvement in several pathways that is found in fibroblasts. Thus cell culture studies were conducted on these cells.

Our cell culture studies revealed that at 250 μM , cellular toxicity was observed for both formulations ($P < 0.05$). At the level of 100 μM , a significant difference occurred between SLN and liposome treated groups. In the SLN exposed group the vitality of the cells were over 80%, however for liposome treated group, dramatic cell death was observed (Figure 4). The critical concentration of Q10 in the nanosystems that the cells can survive was determined as 50 μM for both formulations. At this concentration, in the liposome-treated group, cell proliferation was observed (viability over 120%). At the lowest Q10 concentration of 10 μM , no difference could be seen in terms of cell viability between SLNs and liposome treated groups. However at 25 μM the viability results were again in favor of liposomes (Figure 4). Thus it was decided to carry out additional cell

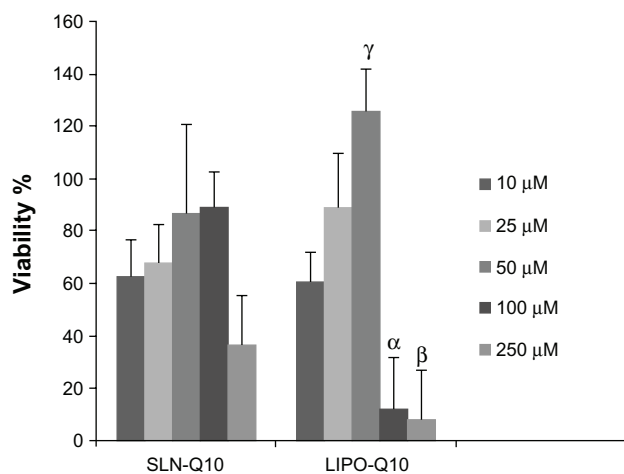


Figure 4 Viability (%) of fibroblast cells after 24 hours of treatment with 10, 25, 50, 100, and 250 μM of SLNs and liposomes.

Notes: The results are the means \pm SD. $\alpha, \beta, \gamma, P < 0.05$; viability % of fibroblasts; SLNs vs liposomes.

Abbreviations: SLN, solid lipid nanoparticle; SD, standard deviation; SLN-Q10, Q10-loaded solid lipid nanoparticles; LIPO-Q10, Q10-loaded liposomes.

viability studies under oxidative conditions at the concentration levels of 25 μM and 50 μM.

Under the oxidative conditions, it was seen that both 25 μM and 50 μM of Q10 concentrations were not able to protect cells from oxidative damage when encapsulated in SLNs. Liposomes performed better protective effects and 50 μM was a better choice of concentration for this aim. The proliferation of the cells led to a viability value, such as 120%. SLN's protective effect was lower when compared to Q10 alone (Figure 5). To understand the underlying mechanism of this result, cytofluorometry studies were conducted at 50 μM.

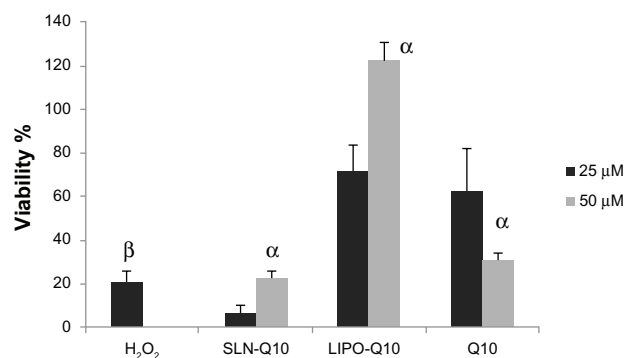


Figure 5 Viability (%) of fibroblast cells treated with 25 μM and 50 μM of SLNs and liposomes after contact with H₂O₂ at concentration of 2 mM.

Notes: The results are the means \pm SD. $\alpha, P < 0.05$; viability % of fibroblasts determined for 50 μM vs 25 μM; $\beta, P < 0.05$; viability % of fibroblasts treated with only H₂O₂.

Abbreviations: SLN, solid lipid nanoparticle; SD, standard deviations; SLN-Q10, Q10-loaded solid lipid nanoparticles; LIPO-Q10, Q10-loaded liposomes; Q10, coenzyme Q10.

The number of cells versus fluorimetric intensity is presented in Figure 6. 1 mM of DCFH-DA was used to differentiate ROS fluorescence and noise. ROS production in the cells comes as an output of fluorescence. In the case of this study, if the fluorescence intensity gets higher, it means that the antioxidant effect of the tested formulation is lower.

Figure 7 presents the histograms of cells versus fluorimetric intensity evaluated for Q10, SLN-Q10, LIPO-Q10, control, and H₂O₂ treated samples. The viability of cell substrates treated with SLN and liposome samples was confirmed with the morphologic parameters (Figure 7), to indicate that the oxidative damage did not cause cell death.

The oxidative damage caused by H₂O₂ treatment is directly related fluorescence intensity due to ROS production. The fluorescence intensity gets higher as the intracellular ROS concentration gets higher. The highest ROS accumulation was determined with H₂O₂ treated cell substrate, as indicated by cell peak characterized by fluorescence intensity ranging from 10 to 100. The mean fluorescence intensity % result obtained from untreated cell substrate was 39.55%. The results obtained from formulation treated and H₂O₂ treated groups could be listed in order as:

$$\text{H}_2\text{O}_2_{(49.27)} > \text{Q10}_{(44.86)} \geq \text{SLN-Q10}_{(44.34)} > \text{LIPO-Q10}_{(27.41)}$$

In the peroxide-treated cells the fluorescence reaches an order of magnitude higher than the control to indicate a higher oxidative damage in absence of an antioxidant. The liposome-treated cell substrate was characterized by a significantly low ROS accumulation. The cell substrates treated with SLN-Q10 were characterized by cell fluorescence intensity almost similar to the substrate treated with only Q10. This result was surprising because of the expected advantage of SLNs as an enhancer in cellular uptake due to nanosize. In our previous study, resveratrol (RSV)-loaded NLC showed fluorescence below that of RSV and RSV-loaded SLN, indicating less ROS production, and this result was attributed to smaller dimensions of RSV-loaded NLC with a reduced negative electrical charge.¹⁰ However, in this study, the dimension of LIPO-Q10 was nearly 2-fold that of SLN-Q10 with the magnitude of ZP higher than SLN-Q10.

It was thought that, since Q10 was highly lipophilic (practically insoluble in water), a delayed release might be responsible for this result. In the studies of Farboud et al³⁰ and Teeranachaideekul et al⁶ it was reported that Q10 was released slowly from the solid matrix of the lipid. In another study, no Q10 release could be seen during 7 days when

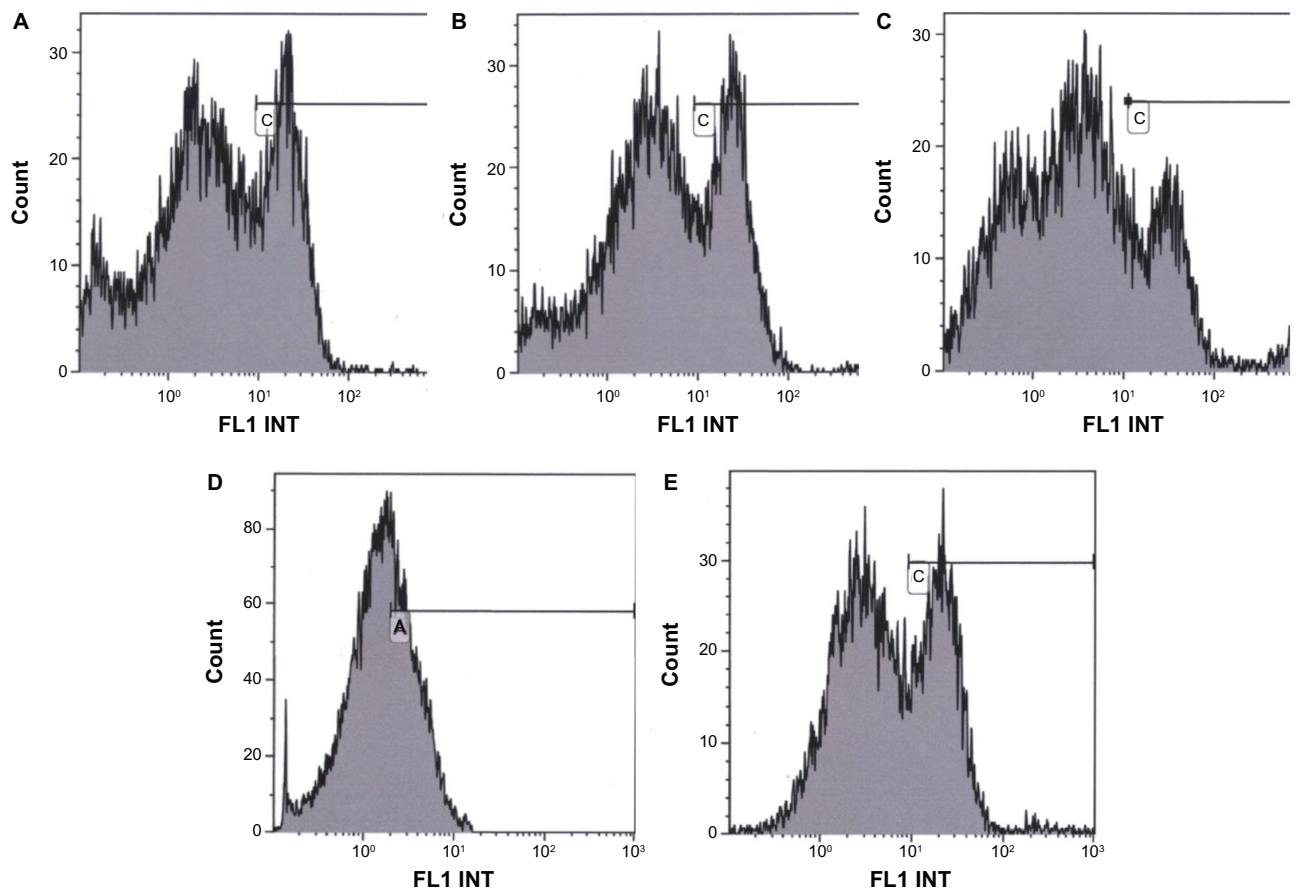


Figure 6 Signal intensity histograms of cells incubated with (A) Q10, (B) SLN-Q10, and (C) LIPO-Q10; (D) control; (E) H₂O₂-treated samples.
Abbreviations: FL1 INT, fluorimetric intensity; Q10, coenzyme Q10; SLN-Q10, Q10-loaded solid lipid nanoparticles; LIPO-Q10, Q10-loaded liposomes.

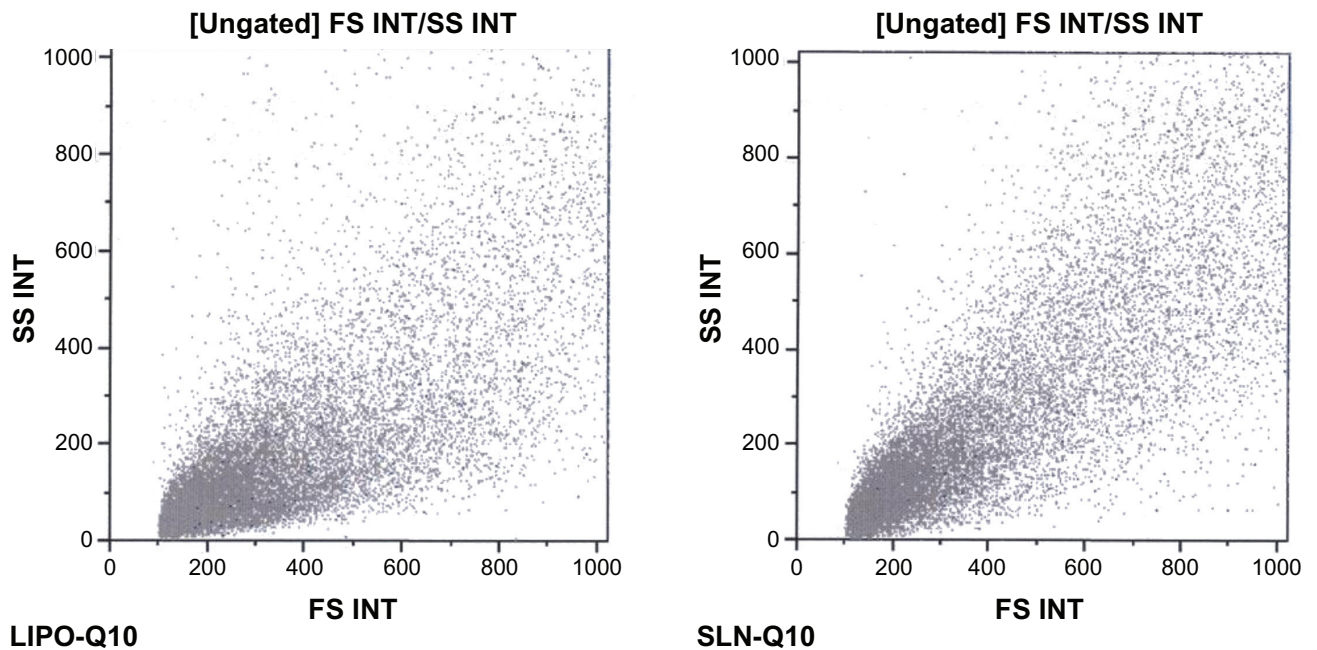


Figure 7 Morphologic parameter of fibroblasts treated with SLN and liposome samples.
Abbreviations: SLN, solid lipid nanoparticle; SLN-Q10, Q10-loaded solid lipid nanoparticles; LIPO-Q10, Q10-loaded liposomes; FS INT, forward scatter; SS INT, side scatter.

the nanoparticles were suspended in water.³¹ On the other hand, liposomes may interact with the cell membranes and disorder the membrane properties.³² It was shown that, the permeability barrier of the stratum corneum was weakened by phosphatidylcholine.³³ Makabi-Panzu et al³⁴ reported that incorporation of Q10 in liposomes enhanced the cellular uptake by macrophage cells. Therefore it was thought that the superior antioxidant effect of LIPO-Q10 observed in this study might be due to membrane perturbing properties of liposomes such as adsorption and fusion.³⁵

Conclusion

Nanosystems made from different structures were previously shown to enhance dermal uptake or improve tolerability of active substances. In this study, SLN-Q10 and LIPO-Q10 were prepared by means of high shear homogenization and thin film hydration method, respectively. The PS of SLNs obtained was significantly smaller than liposomes. However the overall charge was lower in comparison to liposomes. Q10 was not in a crystalline state for both of the nanosystems, indicating its solubility in lipid phases.

After cell culture tests conducted under normal and oxidative conditions, 50 μM was considered the efficient Q10 concentration for cell viability. LIPO-Q10 performed better cell proliferation-inducing properties and ROS production inside the cells was decreased by nearly 50% in comparison to the negative control.

As a conclusion, in terms of dermal antioxidant activity of Q10, liposomes were regarded as better and more promising delivery systems in comparison to SLNs.

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Disclosure

The authors report no conflicts of interest in this work.

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